Journal of Organometallic Chemistry, 197 (1980) 39-43 Elsevier Sequoia, S.A., Lausanne — Printed in The Netherlands

2'3'-ISOPROPYLIDENE 6-MERCAPTOPURINE RIBOSIDE(S-,N-)TRI-n-BUTYLTIN

GEORGE DOMAZETIS *, ROBERT J. MAGEE and BRUCE D. JAMES

Department of Inorganic and Analytical Chemistry, La Trobe University, Bundoora, Vic. 3083 (Australia)

(Received March 18th, 1980)

Summary

The synthesis and characterization of 2'3'-isopropylidene 6-mercaptopurine riboside(S-,N-)tri-n-butyltin is presented. Spectroscopic data indicate that the complex is five-coordinate with an Sn—S bond and N(7) coordination to tin. IR and 1H NMR spectra indicate association via CH_2OH --- N(1) intermolecular H-bonding.

Heterocyclic ligands such as 2-mercaptobenzothiazole and 2-mercapto-4,6-hydroxy-pyrimidine have been shown to react readily with triorganotins under mild conditions and the ligands may bind as either the thiol or the thione form [1]. No identifiable products however, were obtained when thiouracil or 2-amino-6-mercaptopurine were allowed to react under similar conditions. Impure products could be obtained in boiling ethanol or by refluxing in toluene [2].

Our interest in heterocyclic ligands stems from (i) a need to evaluate the influence of tautomerism on the reactivity of -SH and -NH groups towards organotins, and (ii) the likelihood that organotins may bind to S,N groups in biological systems. The ligand 2'3'-isopropylidene-6-mercaptopurine riboside is useful for studies relating to these aspects. A triorganotin may form either a four-coordinate complex through bonding to sulfur or a five-coordinate complex through additional N or O coordination. Sulfur and N-7 coordination by 6-mercaptopurines has been observed in a number of complexes [4,5]. We report here the synthesis and characterization of the first trioganotin derivative of 2'3'-isopropylidene-6-mercaptoriboside, and propose a structure based on NMR, IR and UV spectra.

^{*} Present address: Department of Chemistry, University of British Columbia, Vancouver, B.C., V6T 1Y6 (Canada).

Experimental

The complex was prepared by adding 0.67 g (2.06 mmol) of 2'3'-isopropylidene 6-mercaptopurine riboside (Aldrich) to 30 cm³ CHCl₃ followed by 0.62 g (1.03 mmol) bis(tributyltin) oxide (Alfa). The mixture was stirred for 1—3 h. The solvent was evaporated off in an air stream, the product dissolved in 50 cm³ methanol and stirred for ca. 1 h after which the methanol was air evaporated. The thick, clear oil was dried in a vacuum desiccator for a week. Analysis Found: C, 48. 67; H, 7.11; N, 8.2; S, 5.1; Sn 20.1, C₂₅H₄₂O₄H₄SSn Calcd.: C, 48.96; H, 6.89; N, 9.14; S, 5.23; Sn, 19.36%. The yield was essentially quantitative. The product appears to be thermally stable at room temperature and is soluble in all common organic solvents.

Infrared spectra were obtained using a Perkin-Elmer 457 instrument calibrated with polystyrene film and UV spectra were obtained from CHCl₃ solutions with a Varian 634 instrument.

The ¹H spectra were recorded at ambient temperature using a 90 MHz Perkin-Elmer R32 instrument. For ¹³C NMR spectra, the ligand was dissolved in DMSO- d_6 and the complex in CDCl₃ and the spectra were obtained using a JEOL-PFT-100FT spectrometer as described previously [3].

Results and discussion

The compound has been characterized by chemical analyses, 13 C and 1 H NMR, IR and UV spectroscopy. A n-Bu₃Sn-S complex with nitrogen coordination is indicated by the 13 C NMR spectrum. The α C chemical shift may be compared to that in other compounds containing similar bonding, and the ^{1}J value is also similar (See Table 1).

In the IR spectrum of the complex, the broad peaks observed in the 3000—2600 cm⁻¹ region for the ligand are no longer present. This shows loss of the SH or the NH proton from the tautomers:

Bands at 590 cm⁻¹ and 515 cm⁻¹ are predominantly $\nu(Sn-C)$ with some overlapping from ligand peaks. The bands which have shifted relative to those of the free ligand are shown in Table 2 (not all bands listed).

The vibrational modes associated with the purine ring (and the ribose portion) are shifted on compound formation. The shift in $\nu(O-H)$ is indicative of strong H-bonding. No evidence for $Sn \leftarrow O$ coordination can be obtained from IR or NMR data. The changes in the purine ring vibrational modes are likely to be the result of mass effects from $Sn \leftarrow N$ coordination. The changes

TABLE 1

13C NMR RESULTS

Compound	aC Shift a	¹ J(¹¹⁹ Sn- ¹³ C)	2J	3 _J
Title complex	16.1	343	21	_
n-Bu ₃ Sn(2-mercaptobenzothiazole)	16.4	336	22	62
n-Bu ₃ Sn(2-thiol 5-nitropyridine)	16.3	356	22	78
n-Bu ₃ Sn(2-thiol pyridine oxide)	19.0	361	_	63
Other shift values $\beta C = 28.8$, $\gamma C = 27.0$, $\delta C = 13.7$	(i) Organotin group:			
20.0, 10 21.0, 00 10.1	(ii) Ligand:			
166.6, 150.0, 146.9, 142.0 (134.8?), 11 81.7, 63.2, (28.3?), 27.5, 25.2	** * */	3.5		

a Shift values are in ppm relative to internal TMS.

associated with the ribose portion may result from stronger H-bonding involving the -CH₂OH group. A band at 638 cm⁻¹ is assigned (tentatively) to $\nu(Sn \leftarrow N)$ [8].

The ¹H NMR spectra of the complex and ligand (Fig. 1) differ significantly. The shifts to higher field of the C-2 and C-8 protons indicate nitrogen coordination. Changes in the riboside proton chemical shifts, and the concentration dependence of all proton chemical shifts, suggests intermolecular association via H-bonding. The UV spectra of ligand and complex are consistent with Si \leftarrow N coordination. The ligand band centred at 340 nm (broad) shifts on compound formation and is present as a narrow strong band at 310 nm with a shoulder at 303 nm. These changes are similar to those observed between the UV spectra of 2-thiol-5-nitro pyridine and its di-n-butyltin complex which contains Sn-S and Sn \leftarrow N bonding [9].

The figure below shows the changes in the C-2 and C-8 protons both on compound formation and on dilution. The shift to higher field is probably due to

TABLE 2 INFRARED SPECTRA

Ligand (cm ⁻¹)	Complex (cm ⁻¹)	Assignment	Reference
3400m(br)	3290m(br)	ν(O-H)	
1612s, 1593s	1568vs	vibr. of purine	6, 7
1583s			
1417s	1425s		
1430w			
1209s	1220(sh), 1203s		
1155m	1142m		
1100(sh), 1089s	1100s }	degenerate vibr.	6
1072s, 1055s		of C—C—C in ribose	
		О	
880m, 868m	875(sh)		
663w, 652m	670m, 650m	ribose system degenerate vib.	6
	638m	ν(SnN) (?)	8

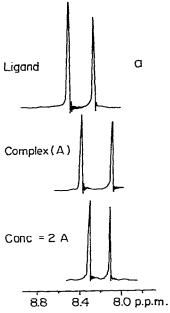
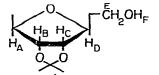


Fig. 1. ¹H NMR of the complex and the ligand.

TABLE 3 ^{1}H NMR SPECTRA OF COMPLEX AND LIGAND

Chem. shift ^a	Peak pattern	Signal sep. (Hz)	No. of protons	Assignment
Complex				
5.91	doublet	$^{1}J = 3.2$	1	A
5.81-5.91	quartet	3.2, ca. 9	1	fB+C+D
5.11	4-peaks	_	2	· ·
4.49	broad, single	_	1	E
3.89	broad, 4 peaks		2	F
1.63	single	_	3	CH ₃
8.35	single	_	1	C-2
8.03	single	-	1	C-8
Ligand				
6.14	doublet	$^2J=2.8$	1	A
5.34, 5.27	two doublets	2,8, ca. 6	1	B (?)
5.09	broad triplet	ca. 4	1	F
5.00, 4.93	two doublets	2.8, ca. 6	1	C (?)
4.28	six peaks	_	1	D (?)
3.57	broad triplet	ca. 4	2	E
3.32	broad single	-	3	(N—H, S—H?)
1.55	single	_	3	∫CH3
1.34	single	_	3	[≀] CH ₃
8.49	single		1	C-2
8.24	single		1.	C-8

 $^{^{}a}$ ppm relative to TMS.



coordination by N-7 to tin, while the change on dilution (downfield for C-2 proton) may result from a disruption of H-bonding involving N-1.

The spectra of the riboside portion has changed on compound formation; the values are compared in Table 3. Vicinal coupling constants are sensitive to changes in the dihedral angle [10], while changes in the apparent chemical shifts may arise from a change in the relative orientation of the purine ring. An analysis of the four-spin system, and a study of the solvent-concentration effects is necessary before quantitative deductions may be drawn. The data

presented is consistent with the structure shown: a five-membered chelate ring involving N-7 to tin coordination, and H-bonding between N-1 and the hydroxyl group.

Acknowledgement

G.D. acknowledges support from a Commonwealth of Australia Postgraduate Research Award.

References

- 1 G. Domazetis, Ph.D. Thesis, La Trobe University, 1979.
- 2 T. Ogawa and M. Tatsui, J. Organometal. Chem., 145 (1978) C37.
- 3 G. Domazetis, R.J. Magee and B.D. James, J. Organometal, Chem., 148 (1978) 339.
- 4 E. Sletten, in B. Pullman and N. Goldblum (Eds.), Metal-Ligand Interactions in Organic Chemistry and Biochemistry, part 1, D. Reidel Publishing Co., Dordrecht, 1977, pp. 53-64.
- 5 N. Kottmair and W. Beck, Inorg. Chim. Acta, 34 (1979) 137. M.J. McCall and M.R. Taylor, Acta Cryst. B, 32 (1976) 1687. P. DcMeester, D.M.L. Goodgame, A.C. Skapski and Z. Warnke, Biochem. Biophys. Acta, 324 (1973) 301.
- 6 T. Shimanouchi, M. Tsuboi and Y. Kyogoku, Adv. Chem. Phys., 7 (1964) 435.
- 7 C.L. Angell, J. Chem. Soc., 504 (1961).
- 8 A. Marchand, M. Riviere Baudet, R. Gassend and M.H. Soulard, J. Organometal. Chem., 118 (1976)
- 9 G. Domazetis, B.D. James, M.F. Mackay and R.J. Magee, J. Inorg. Nucl. Chem., 41 (1979) 1555.
- 10 K. Wuthrich, NMR in Biological Research: Peptides and Proteins, North-Holland Publishing Co., Amsterdam, 1976, p. 44.